

## H.pylori Antigen Rapid Test Cassette (Feces)

### INTENDED USE fessional in vitro diagnostic use only

specimens to aid in the diagnosis of H.pylori infection The H.pylori Antigen Rapid Test Cassette (Feces) is a rapid immunoassay for the qualitative detection of H.pylori antigens in human feces

### (SUMMARY)

and costly invasive diagnostic methods include gastric or duodenal biopsy followed by urease testing (presumptive), culture, and/or histologic staining. A very common approach to the diagnosts of *H.pylori* infection is the serological identification of specific antibodies in infected patients. The main limitation of serology test is the inability to distinguish current and past infections. Antibody may be present in the patient's serum long after eradication of the organisms. HpSA (H. pylori Stool Antigen) testing is gaining popularity for diagnosis of H. pylori infection and also for monitoring the efficacy of the treatment of H. pylori infection. Studies have found that more than 90% of patients with duodenal ulcer and 80% of patients with gastric ulcer are infected with H. pyloris. with H.pylori infection in patients with symptoms of gastrointestinal disease. Specimen-dependent including duodenal and gastric ulcer, non-ulcer dyspepsia and active and chronic gastritis. 1,2 Both invasive and non-invasive methods are used to diagnose *H.pylori* H.pylori is a small, spiral-shaped bacterium that lives in the surface of the stomach and duodenum. It is implicated in the etiology of a variety of gastrointestinal diseases,

The *H.pylori* Antigen Rapid Test Cassette (Feces) is a rapid chromatographic immunoassay for the qualitative detection of *H.pylori* antigens in human feces specimens, providing results in 10 minutes. The test utilizes antibodies specific for *H.* pylori antigens to selectively detect H.pylori antigens in human feces specimens.
PRINCIPLE

will always appear in the control line region indicating that proper volume of specimen has been added and membrane wicking has occurred. with anti-H.pylori antibodies on the membrane and generate a colored line. The presence of this colored line in the test region indicates a positive result, while its absence indicates a negative result. To serve as a procedural control, a colored line immunoassay for the detection of *H.pylori* antigens in human feces specimens. In this test, the membrane is pre-coated with anti-*H.pylori* antibodies on the test line region of the test. During testing, the specimen reacts with the particle coated with anti-*H.pylori* the test. During testing, the specimen reacts with the particle coated with anti- $H_{\nu}$ pylon antibodies. The mixture migrates upward on the membrane by capillary action to react The H.pylori Antigen Rapid Test Cassette (Feces) is a ateral

### [REAGENTS]

The test cassette contains monoclonal anti-H.pylori antibodies coated nal anti-H.pylori antibodies coated on the membrane. particles

### [PRECAUTIONS]

- For professional in vitro diagnostic use only. Do not use after expiration date
- The test should remain in the sealed pouch until use.
- the standard procedures for proper disposal of specimens. precautions against microbiological hazards throughout all procedures and follow Handle all specimens as if they contain infectious agents. Observe established Do not eat, drink or smoke in the area where the specimens or kits are handled
- protection when specimens are assayed.

  The used test should be discarded according to local regulations. Wear protective clothing such as laboratory coats, disposable gloves and
- Humidity and temperature can adversely affect results

## (STORAGE AND STABILITY)

The kit can be stored at room temperature or refrigerated (2-30°C). The test cassette must stable through the expiration date printed on the sealed pouch. The test cassette must remain in the sealed pouch until use. **DO NOT FREEZE**. Do not use beyond the

# [SPECIMEN COLLECTION AND PREPARATION]

- no detergents, preservatives or transport media The feces specimen must be collected in clean, dry, waterproof container containing
- Bring the necessary reagents to room temperature before use. If specimens are to be shipped, they should be packed in compliance with federal egulations covering the transportation of etiologic agents

### [ MATERIALS

Package insert

Specimen collection tubes with extraction buffer **Materials Provided** 

Specimen collection containers Materials Required But Not Provided Pipette and disposable tips (optional)
 Droppers

### DIRECTIONS FOR USE Timer

specimen, buffer and/or controls ö reach room temperature

Collect sufficient quantity of feces (1-2 mL or 1-2 g) in a clean, dry specimen collection container to obtain maximum antigens (if present). Best results will be obtained if the assay is performed within 6 hours after collection. Specimen collected may be stored for 3 days at 2-8°C if not tested within 6 hours. For long term storage, specimens should be kept below ~20°C. 5-30°C) prior to testing. To collect fecal specime

To process fecal specimens:

<u>For Solid Specimens:</u>

<u>For Solid Specimens:</u>

Unscrew the cap of the specimen collection tube, then randomly stab the specimen collection applicator into the fecal specimen in at least 3 different sites to collect approximately 50 mg of feces (equivalent to 1/4 of a pea). Do not scoop the fecal specimen.

For Liquid Specimens:
 Hold the dropper vertically, aspirate fecal specimens, and then transfer 2 drops (approximately 80 µL) into the specimen collection tube containing the extraction buffer.

Tighten the cap onto the specimen collection tube, collection tube vigorously to mix the specimen and the then shake the extraction buffer. specimenLeave the

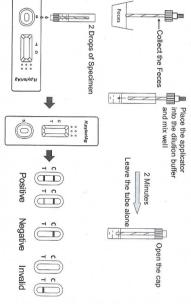
tube alone for 2minnutes

3. Bring the pouch to room temperature before opening it. Remove the test cassette from the foil pouch and use if within one hour. Best results will be obtained if the test is performed immediately after opening the foil pouch.

Hold the specimen collection tube upright and open the cap onto the specimen collection tube. Invert the specimen collection tube ube. Invert the specimen collection tube and transfer 2 full drops of the extracted specimen (approximately 80 µL) to the specimen well (\$) of the test cassette, then start the timer. Avoid trapping air bubbles in the specimen well (\$).

Read results at 10 minutes after dispensing the specimen. Do not read results after 20 minutes.

Note: If the specimen does not migrate (pre-extracted specimens contained in the extract supernatant, dispense into the specimen well (s afresh following the instructions mentioned above: αν γνιενειτίε or particles), centrifuge the extraction buffer vial. Collect 80 μL of the vell (S) of a new test cassette and start shove



[INTERPRETATION OF RESULTS]

(Please refer to the illustration above)

POSITIVE.\* Two lines appear. One colored line should be in the control line region (C) and another apparent colored line should be in the test line region (T).

\*NOTE: The intensity of the color in the test line region (T) will vary depending on the concentration of H.plyfor antigen present in the specimen. Therefore, any shade of color in the test line region (T) should be considered positive.

NEGATIVE: One colored line appears in the control line region (C). No line appears the test line region (T).

INVALID: Control line fails to appear, Insufficient specimen volume or incorrect procedural techniques are the most likely reasons for control line failure. Review the procedure and repeat the test with a new test. If the problem persists, discontinuusing the test kill immediately and contact your local distributor.

[QUALITY CONTROL] e. Review the s, discontinue

Internal procedural controls are included in the test. A colored line appearing in the control region (C) is an internal valid procedural control. It confirms sufficient specimen volume and correct procedural technique.

Control standards are not supplied with this kit, however, it is recommended that positive and negative controls be tested as a good laboratory practice to confirm the test procedure and to verify proper test performance.

### [LIMITATIONS]

1. The *H.pylori* Antigen Test Cassette (Feces) is for in vitro diagnostic use only. The test should be used for the detection of *H.pylori* antigens in feces specimens only. Neither the quantitative value nor the rate of increase in *H.pylori* antigens concentration can be determined by this qualitative test.

2. The *H.pylori* Antigen Test Cassette (Feces) will only indicate the presence of *H.pylori* in the specimen and should not be used as the sole criteria for *H.pylori* to be etiological agent for peptic or duodenal ulcer.

3. As with all diagnostic tests, all results must be interpreted together with other clinical information available to the physician.

information available to the physician.

4. If the test result is negative and clinical symptoms persist, additional testing using other clinical methods is recommended. A negative result does not at any time preclude the possibility of *H.pylori* infection.

5. Following certain antibiotic treatments, the concentration of *H.pylori* antigens may decrease to the concentration below the minimum detection level of the test. Therefore, diagnosis should be made with caution during antibiotic treatment.

**[EXPECTED VALUES]**The *H.pylori* Antigen Test Cassette (Feces) has been compared with Endoscope based methods, demonstrating an overall accuracy of 99.1%.

## PERFORMANCE CHARACTERISTICS,

The *H.pylori* Antigen Test Cassette (Feces) has been evaluated with specimens obtained from a population of symptomatic and asymptomatic individuals. The result shows that the sensitivity of the *H.pylori* Antigen Test Cassette (Feces) is >99.9% and the specificity is 98.4% relative to Endoscope-based methods.

0.21	_	Cassette (Feces) Positive 100 2	H nylori Antigen Test Results Positive Negative	Method method
021	100	0 2		method
	420	102		Total Result

Relative Sensitivity: >99.9% (95%CI\*: 97.0%-100%) \*Confidence Interval

Relative Specificity: 98.4% (95%CI\*: 94.2%-99.8%) Accuracy: 99.1% (95%CI\*; 96.8%-99.9%)

Intra-Assay
Within-run precision has been determined by using 15 inegative, low titer positive, middle titer positive and high specimens were correctly identified >99% of the time. replicates of four specimens titer positive specimens. The

Inter-Assay

Between-run precision has been determined by 15 independent assays on the same four specimens: negative, low titer positive middle titer positive and high titer positive specimens. Three different lots of the H.pylori Antigen Test Cassatte (Feces) have been tested using these specimens. The specimens were correctly identified >99% of the H.pylori Antigen Test Cassatte (Feces) have been tested using these specimens. The specimens were correctly identified >99% of the H.pylori Antigen Tested using these specimens. the time

ollowing organisms were found negative when Cross-reactivity studied at 1.0E+09 organisms/ml. n tested with the *H.pylori* Antigen

Group A Streptococcus
Hemophilus influenza The following organisms were Test Cassette (Feces): Acinetobacter calcoaceticus Av Candida albicans Acinetobacter spp Chlamydia trachomatis Enterococcus faecalis Group B Streptococcus Klebsiella pneumonia Proteus mirabilis Branhamella catarrhalis
Enterococus faecium
Gardnerella vaginalis
Group C Streptococcus
Neisseria gonorrhea
Proteus vulgaris
Salmonella choleraesius

### Neisseria meningitides Pseudomonas aeruginosa Staphylococcus aureus 【BIBLIOGRAPHY】 Adenovirus

Marshall, all, BJ, McGechie, DB, Rogers, PAR ylobacterinfection and gastroduodenal disease. and Glancy, RG. Pyloric Med. J. Australia. (1985), 149

markerof bacte 82(4): 292-96 4. 4 Cutler AF, 1 Campylobacternnection and you will implications for therapy. New England J. 39-44.
Soil, AH. Pathogenesis of peptic ulcer and implications for therapy. New England J. Soil, AH. Pathogenesis of peptic ulcer and implications for therapy. New England J. Soil, AH. Pathogenesis of peptic ulcer and implications for therapy. New England J. Soil, AH. Pathogenesis of peptic ulcer and implications for therapy. New England J. Soil, AH. Pathogenesis of peptic ulcer and implications for therapy. New England J. Soil, AH. Pathogenesis of peptic ulcer and implications for therapy. New England J. Soil, AH. Pathogenesis of peptic ulcer and implications for therapy. New England J. Soil, AH. Pathogenesis of peptic ulcer and implications for therapy. New England J. Soil, AH. Pathogenesis of peptic ulcer and implications for therapy. New England J. Soil, AH. Pathogenesis of peptic ulcer and implications for therapy. New England J. Soil, AH. Pathogenesis of peptic ulcer and implications for therapy. New England J. Soil, AH. Pathogenesis of peptic ulcer and implications for therapy. New England J. Soil, AH. Pathogenesis of peptic ulcer and implications for therapy. New England J. Soil, AH. Pathogenesis of peptic ulcer and implications for the peptic ulcer. All the peptic ulcer. A

100:35S-41S. 5. Anand BS, Raed AK, N normalindividual with 1996,91:1112-1115. Testing for Helicobacter pylori in clinical practice. Am j. Med. 1996. Malaty HM, et al. Loe Helicobacter pylori point prevalence of peptic ulcer in infection. Am J Gastroenterol.

	Index of Symbols	Symbols		
Attention, see instructions for use	$\langle \Sigma \rangle$	Tests per kit	EC REP	<sub>ω</sub>
For in vitro		Use by	<b>⊗</b>	
Store between 2-30°C	TOT	Lot Number	REF	
Do not use if package is				
domona				

S

lumber	Use by	ests per kit
REF	<b>⊗</b>	EC REP
Catalog #	Do not reuse	Authorized Representative

Hangzhou AllTest Biotech Co., Ltd. #550, Yinhai Street Hangzhou Economic & Technological Development Area Hangzhou - 310018, P. R. China



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Effective date: 2016-03-14 145019506